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October 14, 2004

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APPLICATION NUMBER: 10/663,955

FILING DATE: September 16, 2003

RELATED PCT APPLICATION NUMBER: PCT/US04/30263

Certified by



Jon W Dudas

Acting Under Secretary of Commerce
for Intellectual Property
and Acting Director of the U.S.
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UTILITY PATENT APPLICATION TRANSMITTAL <i>(Only for new nonprovisional applications under 37 CFR 1.53(b))</i>	Attorney Docket No.	
	First Inventor	Robert H. Wohleb
	Title	Direct Vial SSMD & Method
	Express Mail Label No.	ER 216 164 183 US

 22278 U.S. PTO
 10/663955
 09/16/03

APPLICATION ELEMENTS See MPEP chapter 600 concerning utility patent application contents.	ADDRESS TO: Mail Stop Patent Application Commissioner for Patents P.O. Box 1450 Alexandria VA 22313-1450
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1. ☒ Fee Transmittal Form (e.g., PTO/SB/17)
(Submit an original and a duplicate for fee processing)
2. ☒ Applicant claims small entity status.
See 37 CFR 1.27.
3. ☒ Specification [Total Pages 18]
(preferred arrangement set forth below)
 - Descriptive title of the invention
 - Cross Reference to Related Applications
 - Statement Regarding Fed sponsored R & D
 - Reference to sequence listing, a table, or a computer program listing appendix
 - Background of the Invention
 - Brief Summary of the Invention
 - Brief Description of the Drawings (if filed)
 - Detailed Description
 - Claim(s)
 - Abstract of the Disclosure
4. ☒ Drawing(s) (35 U.S.C. 113) [Total Sheets 4]
5. Oath or Declaration [Total Sheets 2]
 - a. ☒ Newly executed (original or copy)
 - b. ☐ Copy from a prior application (37 CFR 1.63(d))
(for continuation/divisional with Box 18 completed)
 - i. ☐ **DELETION OF INVENTOR(S)**
Signed statement attached deleting inventor(s) name in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b).
6. ☐ Application Data Sheet. See 37 CFR 1.76

7. ☐ CD-ROM or CD-R in duplicate, large table or Computer Program (Appendix)
8. Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)
 - a. ☐ Computer Readable Form (CRF)
 - b. Specification Sequence Listing on:
 - i. ☐ CD-ROM or CD-R (2 copies); or
 - ii. ☐ Paper
 - c. ☐ Statements verifying identity of above copies

ACCOMPANYING APPLICATION PARTS

9. ☒ Assignment Papers (cover sheet & document(s))
10. ☐ 37 CFR 3.73(b) Statement (when there is an assignee) ☒ Power of Attorney
11. ☐ English Translation Document (if applicable)
12. ☒ Information Disclosure Statement (IDS)/PTO-1449 ☒ Copies of IDS Citations
13. ☐ Preliminary Amendment
14. ☒ Return Receipt Postcard (MPEP 503)
(Should be specifically itemized)
15. ☐ Certified Copy of Priority Document(s)
(if foreign priority is claimed)
16. ☐ Nonpublication Request under 35 U.S.C. 122 (b)(2)(B)(i). Applicant must attach form PTO/SB/35 or its equivalent.
17. ☐ Other:

18. If a CONTINUING APPLICATION, check appropriate box, and supply the requisite information below and in the first sentence of the specification following the title, or in an Application Data Sheet under 37 CFR 1.76:

☐ Continuation ☐ Divisional ☐ Continuation-in-part (CIP) of prior application No.:

Prior application information:

Examiner

Art Unit:

For CONTINUATION OF DIVISIONAL APPS only: The entire disclosure of the prior application, from which an oath or declaration is supplied under Box 5b, is considered a part of the disclosure of the accompanying continuation or divisional application and is hereby incorporated by reference. The incorporation can only be relied upon when a portion has been inadvertently omitted from the submitted application parts.

19. CORRESPONDENCE ADDRESS
☐ Customer Number: OR ☒ Correspondence address below

Name	Keeling Hudson, L.L.C.				
Address	901 North Post Oak Road				
City	Houston	State	TX	Zip Code	77024
Country	USA	Telephone	713-680-1447	Fax	713-680-8567
Name (Print/Type)	James E. Hudson III	Registration No. (Attorney/Agent)	41,081		
Signature	<i>James E. Hudson III</i>	Date	09/16/03		

This collection of information is required by 37 CFR 1.53(b). The information is required to obtain or retain a benefit by the public which is to file (and by the USPTO to process) an application. Confidentiality is governed by 35 U.S.C. 122 and 37 CFR 1.14. This collection is estimated to take 12 minutes to complete, including gathering, preparing, and submitting the completed application form to the USPTO. Time will vary depending upon the individual case. Any comments on the amount of time you require to complete this form and/or suggestions for reducing this burden, should be sent to the Chief Information Officer, U.S. Patent and Trademark Office, U.S. Department of Commerce, P.O. Box 1450, Alexandria, VA 22313-1450. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Mail Stop Patent Application, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450.

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15623 U.S. PAT. & TM. OFF.
09/16/03

PTO/SB/17 (05-03)
Approved for use through 04/30/2003. OMB 0651-0032
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FEE TRANSMITTAL for FY 2003

Effective 01/01/2003. Patent fees are subject to annual revision.

☒ Applicant claims small entity status. See 37 CFR 1.27

TOTAL AMOUNT OF PAYMENT (\$) 433

Complete if Known

Application Number

Filing Date

First Named Inventor

Robert H. Wohleb

Examiner Name

Art Unit

Attorney Docket No.

METHOD OF PAYMENT (check all that apply)

☒ Check ☐ Credit card ☐ Money Order ☐ Other ☐ None

☒ Deposit Account:

Deposit Account Number
Deposit Account Name

11-0307

Keeling Hudson, L.L.C.

The Director is authorized to: (check all that apply)

☐ Charge fee(s) indicated below ☒ Credit any overpayments

☒ Charge any additional fee(s) during the pendency of this application

☐ Charge fee(s) indicated below, except for the filing fee to the above-identified deposit account.

FEE CALCULATION

1. BASIC FILING FEE

Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid
1001 750	2001 375	Utility filing fee	375
1002 330	2002 165	Design filing fee	
1003 520	2003 260	Plant filing fee	
1004 750	2004 375	Reissue filing fee	
1005 160	2005 80	Provisional filing fee	
SUBTOTAL (1)			(\$) 375

2. EXTRA CLAIM FEES FOR UTILITY AND REISSUE

		Extra Claims		Fee from below	Fee Paid
Total Claims	22	-20** =	2	X 9	= 18
Independent Claims	3	-3** =	0	X	=
Multiple Dependent					=

Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description
1202 18	2202 9	Claims in excess of 20
1201 84	2201 42	Independent claims in excess of 3
1203 280	2203 140	Multiple dependent claim, if not paid
1204 84	2204 42	** Reissue independent claims over original patent
1205 18	2205 9	** Reissue claims in excess of 20 and over original patent

SUBTOTAL (2) (\$) 18

**or number previously paid, if greater; For Reissues, see above

FEE CALCULATION (continued)

3. ADDITIONAL FEES

Large Entity Fee Code (\$)	Small Entity Fee Code (\$)	Fee Description	Fee Paid
1051 130	2051 65	Surcharge - late filing fee or oath	
1052 50	2052 25	Surcharge - late provisional filing fee or cover sheet	
1053 130	1053 130	Non-English specification	
1812 2,520	1812 2,520	For filing a request for ex parte reexamination	
1804 920*	1804 920*	Requesting publication of SIR prior to Examiner action	
1805 1,840*	1805 1,840*	Requesting publication of SIR after Examiner action	
1251 110	2251 55	Extension for reply within first month	
1252 410	2252 205	Extension for reply within second month	
1253 930	2253 465	Extension for reply within third month	
1254 1,450	2254 725	Extension for reply within fourth month	
1255 1,970	2255 985	Extension for reply within fifth month	
1401 320	2401 160	Notice of Appeal	
1402 320	2402 160	Filing a brief in support of an appeal	
1403 280	2403 140	Request for oral hearing	
1451 1,510	1451 1,510	Petition to Institute a public use proceeding	
1452 110	2452 55	Petition to revive - unavoidable	
1453 1,300	2453 650	Petition to revive - unintentional	
1501 1,300	2501 650	Utility issue fee (or reissue)	
1502 470	2502 235	Design issue fee	
1503 630	2503 315	Plant issue fee	
1460 130	1460 130	Petitions to the Commissioner	
1807 50	1807 50	Processing fee under 37 CFR 1.17(q)	
1806 180	1806 180	Submission of Information Disclosure Stmt	
8021 40	8021 40	Recording each patent assignment per property (times number of properties)	40
1809 750	2809 375	Filing a submission after final rejection (37 CFR 1.129(a))	
1810 750	2810 375	For each additional invention to be examined (37 CFR 1.129(b))	
1801 750	2801 375	Request for Continued Examination (RCE)	
1802 900	1802 900	Request for expedited examination of a design application	

Other fee (specify)

*Reduced by Basic Filing Fee Paid

SUBTOTAL (3) (\$) 40

SUBMITTED BY

Name (Print/Type)	James E. Hudson III	Registration No. (Attorney/Agent)	41,081	Telephone	713-680-1447
Signature	James E. Hudson III	Date	09/16/03		

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants: Robert H. Wohleb)
Serial No.:) Art Unit:
Filed:) Examiner:
)

For: Direct Vial Surface Sorbent Micro Extraction Device and Method

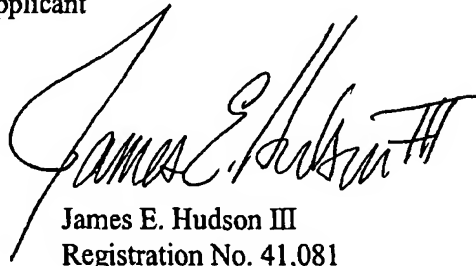
Mail Stop Patent Application
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

September 16, 2003

Dear Sir:

Please find enclosed the following items for Non-Provisional application:

- Return Postcard
- Utility Patent Application Transmittal
- Fee Transmittal
- Check for Filing Fee (\$433.00)
- Declaration and Power of Attorney
- Certificate of Mailing for Declaration and Power of Attorney
- Specification with Claims and Abstract
- Assignment Cover Sheet
- Assignment of Invention
- Four (4) sheets of drawings
- Certificate of Mailing for Drawings
- Information Disclosure Statement
- Information Disclosure Statement by Applicant
- Copies of Listed Information Items



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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant: Robert H. Wohleb

Serial No.:

Filed:

For: Direct Vial Surface Sorbent Micro Extraction Device and Method

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Art Unit:

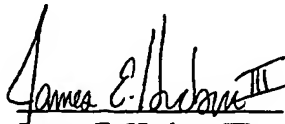
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September 16, 2003

CERTIFICATE OF MAILING

I hereby certify that these four (4) sheets of drawings are being deposited with the United States Postal Service as Express Mail Number ER 216 164 183 US in an envelope addressed to: Mail Stop Patent Application, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450


James E. Hudson III

September 16, 2003
Date of Signature

September 16, 2003
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Houston, Texas 77024-3845
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1 **IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

2 **TITLE OF THE INVENTION**

3 Direct Vial Surface Sorbent Micro Extraction Device and Method.

4 **CROSS-REFERENCE TO RELATED APPLICATIONS**

5 Not Applicable.

6 **STATEMENT REGARDING FEDERALLY SPONSORED**

7 **RESEARCH OR DEVELOPMENT**

8 Not Applicable.

9 **BACKGROUND OF THE INVENTION**

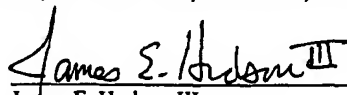
10 Field of the Invention. This invention relates to the extraction and collection of one or
11 more analytes by a sorption process. Specifically, this invention relates to a device and
12 method for performing direct vial extraction.

13 Description of the Related Art. To prepare samples for chemical analysis, often
14 analytes, or the compound of interest, must be separated from a sample matrix, such as water,
15 soil or animal tissue and presented in a form suitable for a particular piece of analytical
16 equipment, such as a gas or liquid chromatograph. There are various extraction methods
17 known and used to collect and prepare samples for such chemical analysis. These methods
18 include liquid/liquid extraction, solid phase extraction, solid phase microextraction and stir-
19 bar sorptive extraction. The trend in the industry is toward simplified sample preparation that
20 results in pollution prevention and waste minimization.

CERTIFICATE OF MAILING UNDER 37 C.F.R. §1.10

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Date: Sept 16, 2003


James E. Hudson III
Registration No. 41,081

1 Liquid/liquid extraction partitions an analyte between two immiscible phases, such as
2 an organic solvent and an aqueous phase. When an aqueous phase contains the analyte it is
3 extracted into the immiscible organic solvent by placing the two phases into contact.
4 Extraction is enhanced by mixing. A relatively large volume of solvent (typically greater
5 than 100 mL) is necessary to carry out the extraction. Partitioning of a compound between
6 the solution solvent and extractant solvent is governed by the distribution constant, K , and
7 the phase ratio, r (The ratio of the quantity of the solvent to that of the other phase). An
8 example of such an extraction would be EPA test method SW846 3510 which specifies that
9 one liter of aqueous sample should be serially extracted with 350 mL of methylene chloride.
10 When the entire procedure is considered, a total of 500 mL of solvent is used for each
11 sample. The solvent extract must be evaporated to reduce its volume to between 1 and 2 mL
12 for placement into an autosampler vial prior to analysis.

13 Solid phase extraction (SPE) is often used to extract a sample prior to analysis by
14 chromatography. SPE uses silica particles with an organic layer covalently attached to the
15 surface of the particles. The silica particles are packed into a tube or disc, such as a
16 polyethylene syringe barrel. The sample is then prepared and an analyte extracted by passing
17 the sample through the solid sorbent. The analyte is then desorbed from the SPE media by
18 solvent extraction. An example of such an extraction is EPA test method SW846 3535 which
19 utilizes one liter of sample but requires approximately 50 mL of solvents. The solvent extract
20 must be evaporated to reduce its volume to between 1 and 2 mL for placement into an
21 autosampler vial prior to analysis.

22 It is known in the art to use a sorbent to extract an analyte from a solution. The

1 analyte is later extracted from the sorbent by thermal desorption or by back extracting with a
2 small amount of organic solvent. Sorption materials are usually homogenous, non-porous
3 materials that are above their glass transition point (T_g) and in which the analyte can dissolve.
4 The sample may be removed for analysis by thermal desorption or solvent extraction.

5 Solid phase microextraction (SPME) is an extraction technique wherein a fiber is
6 coated with a sorbent layer. The coating may be a polysiloxane or other immobilized
7 sorbent. The fiber is immersed in a liquid or exposed to its headspace during which time the
8 analyte is retained. The fiber may then be inserted into a gas chromatograph injection port
9 for analysis where it is thermally desorbed or may be back extracted with a suitable solvent.
10 SPME is not accepted for EPA test methods.

11 Stir-bar sorptive extraction (SBSE) is used primarily for direct mode sampling.
12 SBSE utilizes a thick sorbent coating on a magnetic bar stirrer that stirs the sample for a
13 predetermined amount of time during which time the analyte partitions between the stir-bar
14 sorbent and the sample. After extraction, the stir-bar is removed and the analyte is thermally
15 desorbed to the injection port of a gas chromatograph.

16 Examples of the prior art follow:

17 U.S. Pat. No. 5,595,653 issued to Good et al. on January 21, 1997 discloses an
18 apparatus for extracting an analyte from a liquid sample. The apparatus comprises a
19 microcolumn having a microparticulate media sandwiched between two compression layers.
20 The compression layers are preferably a binder-free glass fiber, held in the microcolumn by
21 upper and lower polypropylene mesh.

22 U.S. Pat. No. 5,635,060 issued to Hagen et al. on June 3, 1997 discloses a solid phase

1 extraction or chromatographic medium. The medium comprises a porous nonwoven fibrous
2 matrix comprising at least one of polytetrafluoroethylene and blown microfibers, and
3 sorptive or reactive hydrophobic siliceous molecular sieve particulates enmeshed in the
4 matrix.

5 U.S. Pat. No. 5,911,883 issued to Anderson on June 15, 1999 discloses a solid phase
6 extraction article having a porous, particle loaded, fibrous sheet material spiral-wrapped
7 around its axis is provided. The sheet material is wound around itself to provide multiple
8 layers of sheet material, each layer of sheet material being spaced from each adjacent layer of
9 sheet material.

10 U.S. Pat. No. 5,897,779 issued to Wisted et al. on April 27, 1999 discloses a cartridge
11 device for removing an analyte from a fluid. The cartridge comprises a hollow core, a sheet
12 composite comprising a particulate-loaded porous membrane and, optionally, at least one
13 reinforcing spacer sheet. The particulate is capable of binding the analyte and the sheet
14 composite is formed into a spiral configuration about the core.

15 U.S. Pat. Nos. 5,415,779 and 5,595,649 both issued to Markell et al. on May 16, 1995
16 and January 21, 1997, respectively, disclose a particle loaded, porous, fibrous compressed or
17 fused article for separations and purifications. The article comprises a nonwoven fibrous
18 polymeric web, which preferably is thermoplastic, melt-extrudable, and pressure-fusible
19 blown microfibrinous web, and sorptive particles enmeshed in the web.

20 U.S. Pat. No. 5,472,600 issued to Ellefson et al. on December 5, 1995 discloses a
21 gradient density filter made from sheets of blown polypropylene microfibers where the
22 microfibers of at least one of the sheets have an effective fiber diameter less than that of the

1 other sheets.

2 U.S. Pat. No. 5,403,489 issued to Hagen et al. on April 4, 1995 discloses a method
3 and apparatus for performing solid phase extraction (SPE) on a fluid that contains solubles
4 and suspended solids. The apparatus includes a conduit, a SPE medium located in the
5 conduit, and a fluid flow direction altering mechanism or a SPE rotating mechanism.

6 U.S. Pat. No. 5,391,298 issued to Pieper et al. on February 21, 1995 discloses an
7 apparatus that can be used to perform a solid phase extraction under pressurized conditions.
8 The apparatus includes a pressurizable housing with an inlet tube that can communicate with
9 a pump, which feeds a liquid to the housing under positive pressure. A disk assembly
10 includes fluid-permeable, porous sheets on opposite sides of an SPE membrane.

11 U.S. Pat. No. 5,279,742 issued to Markel et al. on January 18, 1994, reissued as U.S.
12 Pat. No. Re. 36,811 on August 8, 2000 discloses a method for isolating an environmentally
13 hazardous organic contaminant from a fluid utilizing a solid phase extraction medium. The
14 medium comprises a PTFE fibril matrix, and sorptive particles enmeshed in the matrix. The
15 separations can be efficiently performed in a stacked disk format.

16 U.S. Pat. No. 5,691,206, issued to Pawliszyn on November 25, 1997 discloses a
17 device for carrying out solid phase microextraction. The device is a fiber, solid or hollow,
18 contained in a syringe. The syringe has a barrel, a plunger slidable within the barrel and a
19 hollow needle extending from the end of the barrel opposite the plunger. The needle contains
20 the fiber. When the plunger is depressed, the fiber extends beyond a free end of the needle
21 and when the plunger is in a withdrawn position the fiber is located within the needle. To
22 collect a sample, the needle is inserted through a septum in a bottle containing the sample

1 and the fiber is extended into the sample. After a predetermined amount of time, the fiber is
2 returned to the needle and the syringe is withdrawn from the bottle. The sample is analyzed
3 by inserting the needle through a septum in a gas injection port of a gas chromatograph and
4 extending the fiber.

5 U.S. Pat. No. 5,565,622, issued to Murphy on October 15, 1996 discloses a simplified
6 method for solid phase extraction of components of interest from a sample. A syringe is used
7 in which the inner surface of the cannula or needle is at least partially coated with a
8 stationary phase such that aspirating the sample into the needle results in adsorption of the
9 components of interest into the stationary phase. Aspiration of a solvent may be employed
10 for removing the components of interest from the stationary phase for direct injection into a
11 chromatographic instrument, or the components of interest may be removed by thermal
12 desorption, wherein the needle is placed in the injection port of the chromatographic
13 instrument and heated.

14 U.S. Pat. Application Pub. No. US 2002/0105923, applied for by Malik, published on
15 October 17, 2002 discloses a method of preconcentrating trace analytes by extracting polar
16 and non-polar analytes through a sol-gel coating. The sol-gel coating is either disposed on
17 the inner surface of the capillary tube or disposed within the tube as a monolithic bed.

18 It would be an improvement to the art to have a device in which the extraction may be
19 performed and the analyte conveniently and transportably stored for later analysis.

20

21 BRIEF SUMMARY OF THE INVENTION

22 The present invention comprises a device and method for performing direct vial

1 extraction.

2 Accordingly, the objects of my invention are to provide, inter alia, a single step solid
3 phase extraction system that:

- 4 • minimizes the amount of solvent used;
- 5 • minimizes the amount of labor required to perform an extraction;
- 6 • minimizes glassware;
- 7 • allows samples to be archived;
- 8 • allows extraction to be performed at the sampling site rather than the
9 laboratory;
- 10 • allows the extract to be subjected to replicate analysis;
- 11 • allows the use of gas or liquid chromatography autosamplers;
- 12 • allows the use of disposable sample vials;
- 13 • has greater reproducibility than solid phase micro extraction;
- 14 • reduces or eliminates sample cross contamination; and
- 15 • does not require expensive thermal desorption equipment.

16 This invention is a sorption vial that can be used for the extraction of a sample, or
17 analyte, from a sample matrix and a method of using the sorption vial to perform the
18 extraction. Preferably, the sorption vial has a conically-shaped interior bottom surface coated
19 with sorptive material. An adapter may retain the sorption vial in a fixed position within a
20 larger sample vessel such that the sorptive coating is exposed to a sample or its headspace.
21 After partitioning of the sample in to the sorptive material, the sorption vial may be removed
22 from the sample vessel. An elution solvent is used to extract the analytes from the sorptive

coating, which is then sealed and transported to a location for further testing. Alternatively, the sorption vial may be used directly to receive the sample and perform the extraction without using the larger sample vessel.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a cross-sectional view of a sample vessel with a sorption vial.

Figure 2 is a perspective view of the preferred embodiment of a sorption vial.

Figure 3 is a cross-sectional view of a sorption vial with a vial cap.

Figure 4 is a perspective view of an alternative embodiment of a sorption vial.

DESCRIPTION OF THE INVENTION

Referring to Fig 1, the preferred embodiment of the surface sorbent micro extraction (SSME) assembly is depicted as 10. SSME assembly 10 comprises a sorption vial 20 and a sample vessel 30.

Referring to Figs. 1 and 2, sorption vial 20 is made from a rigid, nonreactive material, such as silica glass. In the preferred embodiment, sorption vial 20 has a cylindrically-shaped interior wall 21 with a conically-shaped bottom surface 22. Sorption vial 20 also has a vial base 40 and a vial neck 26 through which there is an opening 23 to interior surface 22. Bottom surface 22 is oriented such that the vertex 24 of the conical bottom surface 22 is proximate vial base 40 while the directrix 42 is contiguous with interior wall 21.

An alternative embodiment of sorption vial 20 is shown in Fig. 4 as sorption vial 200. Interior wall 222 is conically shaped. Alternative interior wall 222 is oriented such that the vertex 224 of the conical interior wall 222 is proximate vial base 240 while the directrix 242

1 is proximate vial neck 226.

2 It is known in the art that vials need a means for closure. It is also known in the art
3 that autosampers require a means by which they may grasp the vial. Referring to Figs. 2 and
4 3, vial neck 26 is an example of a means known in the art by which vials may be sealed and
5 provide a shape suitable to autosamplers. In this example vial neck is formed such that a vial
6 cap 28 may be placed over opening 23 to seal sorption vial 20 after a sample 15 (shown in
7 Fig. 1) containing the analyte to be extracted is exposed to interior surface 22. Vial cap 28
8 may be any type of cap including a screw-on cap, a crimp cap, or a plug, so long as vial cap
9 28 is leak-proof.

10 A sorptive coating 27 is applied proximate the vertex 24 of interior surface 22. When
11 interior surface 22 is cylindrical rather than conical, sorptive coating 27 may be applied on
12 the cylinder interior wall or the flat or conical bottom surface or both.

13 In the preferred embodiment, the sorptive coating 27 is a hydrophobic coating, such
14 as an immobilized polysiloxane, for example polydimethylsiloxane (PDMS), which contains
15 only methyl functional groups. The name "siloxane" is based on the Si - O - Si unit and has
16 found acceptance in scientific nomenclature. Polysiloxanes are polymers with repeating
17 siloxane units. Each repeating siloxane unit contains two functional groups attached (e.g.
18 dimethyl) which may, or may not, be of the same type of functional group. A functional
19 group is an atom or combination of atoms which gives a polymer its distinctive and
20 characteristic chemistry. A polysiloxane of 50 repeating units would therefore have 100
21 methyl groups, whereas a siloxane unit with two different types of groups such as
22 phenylmethyl would have 50 of each "type" in the polysiloxane.

1 It is known in the art that immobilized polysiloxanes that contain other types of
2 functional groups, may be used as sorbents. These include immobilized polysiloxanes
3 containing phenyl or trifluoropropyl functional groups. Examples of these polysiloxanes
4 include diphenylsiloxane-dimethylsiloxane copolymers and trifluoropropylmethylsiloxanes.
5 For more selective sorption applications the immobilized polysiloxane may contain other
6 types of functional groups including alkyl, alkenyl, alkynyl, aryl, alkylaryl, alkenylaryl,
7 alkynylaryl, haloalkyl or haloaryl. A polysiloxane may contain said types of functional
8 groups in any combination. The selection of the type of functional groups permits the
9 partitioning of a particular analyte or analytes from the sample. The polysiloxane coating may
10 be a polymer, a copolymer or a combination of polymers.

11 Alternatively, sorptive coating 27 may be (1) a porous layer, such as a derivatized
12 etched surface, (2) other immobilized polymers that are above their glass transition
13 temperatures such as poly butadiene, (3) an immobilized porous polymer, such as
14 divinylbenzene, ethyleneglycoldimethacrylate, and copolymers of divinylbenzene and
15 ethyleneglycoldimethacrylate, polyethyleneimine, acrylonitrile, n-vinyl-2-pyrrolidinone or 4-
16 vinyl-pyridine, (4) a sol gel or (5) an immobilized adsorbent such as graphitized carbon
17 black. Sorptive coating 27 may be any one of the coatings described or a combination of
18 two or more of the alternative coatings. The selection of the coating or coatings by one
19 skilled in the art is dependent upon the analyte or analytes to be partitioned from sample.

20 Referring again to Fig. 1, sample vessel 30 is used to collect sample 15 from which
21 the analyte is to be extracted. Sample vessel 30 is made from a rigid, nonreactive material,
22 such as silica glass, and has a mouth 32. A cap 34 is used to close the sample vessel 30 at

1 mouth 32. Cap 34 has an interior surface 35, within which base 40 of sorption vial 20
2 selectively attaches.

3 When sample vessel 30 is closed with sorption vial 20 attached to cap 34, opening 23
4 faces toward sample 15. When sample vessel 30 is sealed and inverted, contained liquid
5 sample 15 contacts sorptive coating 27. Alternatively, sample vessel 30 may be maintained
6 in an upright position with sorption vial 20 exposed to the head space of a collected sample.
7 The analyte within sample 15 is partitioned between sample 15 and sorptive coating 27. The
8 small surface area of interior surface 22 allows for rapid exchange of a vapor or liquid as
9 well as for desorption by the least volume of solvent. Sorption vial 20 may then be removed
10 from cap 34, desorbed by a suitable solvent, sealed and stored or transported from the test
11 collection site to a location for testing.

12 The extraction process comprises placing a sample in sample vessel 30. Sorption vial
13 20 is then attached to cap 34 or cap liner 37 and sample vessel 30 is sealed. As previously
14 explained, sorption vial 20 is attached within sample vessel 30 such that interior surface 22
15 will be exposed to samples within sample vessel 30 or the headspace of such samples.
16 Sample vessel 30 may be agitated for a predetermined period of time to allow equilibrated
17 partitioning. Sorption vial 20 is removed from sample vessel 30. A predetermined amount
18 of elution solvent (not shown) is measured into sorption vial 20, and sorption vial 20 is
19 sealed. The collected sample may be analyzed by gas chromatography, high performance
20 liquid chromatography or other analytical instruments. Alternatively, the collected sample
21 may be stored for future analysis.

22 In certain cases, such as when a sample has a high viscosity, agitation is not desired.

1 In such cases, collection may take place by exposing sorption vial 20 to the headspace of
2 sample 15. Sample vessel 30 may be stirred for a predetermined amount of time to enhance
3 equilibrated partitioning. Partitioning takes place between sample 15, it's headspace and the
4 sorptive coating 27.

5 In some cases the volume of sample is equal to or less than the volume of sorption
6 vial 20. In this case sample vial 20 receives a similar sorptive coating 20 such as PDMS.
7 Sorption vial 20 is then filled with the solution containing analytes to be extracted thus
8 eliminating the need for the sample vessel 30. A mechanical shaker (not shown) is used to
9 agitate sorption vial 20 and to assist in bringing the partitioning to equilibrium. Sorption vial
10 20 is emptied and a predetermined amount of elution solvent (not shown) is measured into
11 sorption vial 20. A vial cap 28 seals sorption vial 20. The contents (not shown) of sorption
12 vial 20 may then be sampled as required. The preferred embodiment of sample vial 20,
13 shown in Fig. 2, is particularly well suited for this method.

14 The foregoing disclosure and description of the invention is illustrative and
15 explanatory thereof. Various changes in the details of the illustrated construction may be
16 made within the scope of the appended claims without departing from the spirit of the
17 invention. The present invention should only be limited by the following claims and their
18 legal equivalents.

CLAIMS

What is claimed is:

1. A device for the collection and extraction of at least one analyte within a sample, said device comprising:

a sorption vial including an interior surface, an opening to said interior surface and a vial base distal said opening;

a sorptive coating on said interior surface;

a sample vessel for collecting said sample;

a cap for closing said sample vessel; and

said vial base selectively attached to said cap.

2. The device of claim 1, further comprising:

said interior surface having a conical shape;

said conically-shaped interior surface having a vertex and a directrix;

said vertex proximate said vial base;

said directrix facing said opening.

3. The device of claim 1, further comprising:

said interior surface including an interior wall and an interior base;

said interior base having a conical shape;

said conically-shaped interior base having a vertex and a directrix;

said vertex proximate said vial base;

said directrix contiguous with said interior wall; and

said sorptive coating covering said conically-shaped interior base.

4. The device of claim 3, wherein said sorption vial comprises silica glass.
5. The device of claim 3, further comprising:
said sorptive coating comprising an immobilized polysiloxane polymer having one type of functional group selected from the group consisting of alkyl, alkenyl, alkynyl, aryl, alkylaryl, alkenylaryl, alkynylaryl, haloalkyl and haloaryl.
6. The device of claim 3, further comprising:
said sorptive coating comprising an immobilized polysiloxane polymer having at least two types of functional groups selected from the group consisting of alkyl, alkenyl, alkynyl, aryl, alkylaryl, alkenylaryl, alkynylaryl, haloalkyl and haloaryl.
7. The device of claim 3, wherein said sorptive coating comprises an immobilized porous polymer.
8. The device of claim 7, wherein said immobilized porous polymer is selected from the group consisting of: divinylbenzene, ethyleneglycoldimethacrylate, polyethyleneimine, acrylonitrile, n-vinyl-2-pyrrolidinone, and 4-vinyl-pyridine.
9. The device of claim 3, wherein said sorptive coating comprises a sol gel coating.
10. The device of claim 3, wherein said sorptive coating is a polymer existing above its glass transition temperature.
11. The device of claim 3, further comprising:
a vial cap for sealing said sorption vial; and
said vial cap covering said opening.
12. A device for the collection and extraction of at least one analyte within a sample, said device comprising:

a sorption vial including an interior wall, an interior base, an opening to said interior wall and said interior base and a vial base distal said opening;

said interior base having a conical shape;

said conically-shaped interior base having a vertex and a directrix;

said vertex proximate said vial base;

said directrix contiguous with said interior wall;

a sorptive coating on said conically-shaped interior base;

a sample vessel for collecting said sample;

a cap for closing said sample vessel;

said vial base selectively attached to said cap;

a vial cap for sealing said sorption vial; and

said vial cap covering said opening.

13. The device of claim 12, further comprising:

said sorptive coating comprising an immobilized polysiloxane polymer having one type of functional group selected from the group consisting of alkyl, alkenyl, alkynyl, aryl, alkylaryl, alkenylaryl, alkynylaryl, haloalkyl and haloaryl.

14. The device of claim 12, further comprising:

said sorptive coating comprising an immobilized polysiloxane polymer having at least two types of functional groups selected from the group consisting of alkyl, alkenyl, alkynyl, aryl, alkylaryl, alkenylaryl, alkynylaryl, haloalkyl and haloaryl.

15. The device of claim 12, wherein said sorptive coating comprises an immobilized porous polymer.

16. The device of claim 15, wherein said immobilized porous polymer is selected from the group consisting of: divinylbenzene, ethyleneglycoldimethacrylate, polyethyleneimine, acrylonitrile, n-vinyl-2-pyrrolidinone, and 4-vinyl-pyridine.
17. The device of claim 12, wherein said sorptive coating comprises a sol gel coating.
18. The device of claim 12, wherein said sorptive coating is a polymer existing above its glass transition temperature.
19. A method for performing direct vial extraction of analytes from a sample utilizing a sorption vial and a sample vessel, said sorption vial including a vial base, a vial interior, and a vial opening, said vial interior intermediate said vial base and said vial opening, said method comprising:
- coating said vial interior with a sorptive material;
 - selectively attaching said vial base to a cap;
 - collecting a liquid sample in said sample vessel;
 - closing said sample vessel with said cap;
 - exposing said liquid sample to said sorptive coating;
 - opening said sample vessel;
 - removing said sorption vial from said cap;
 - adding a solvent to said sorption vial; and
 - sealing said sorption vial with a vial cap.
20. The method of claim 19 wherein said exposing step comprises agitating said sample vessel.

21. The method of claim 19 wherein said exposing step comprises subjecting said sorption vial to a headspace above said sample.
22. The method of claim 19 wherein said vial interior including a conically-shaped bottom surface; and
said coating step including coating said conically-shaped bottom surface with said sorptive coating.

ABSTRACT OF THE INVENTION

A device for extracting an analyte from a sample matrix comprises a sorption vial with a conically shaped interior surface, which is coated with a sorbent material. A method for extracting an analyte from a sample matrix includes retaining the sorption vial within a sample vessel with the sorbent coating exposed to the sample matrix contained in the sample vessel. After the analyte is collected in the sorbent material, the sorption vial may be removed from the sample vessel and sealed, or a small amount of elution solvent may be added to the sorption vial before sealing. The sorption vial containing the analyte may then be stored or transported to a lab for further analysis.

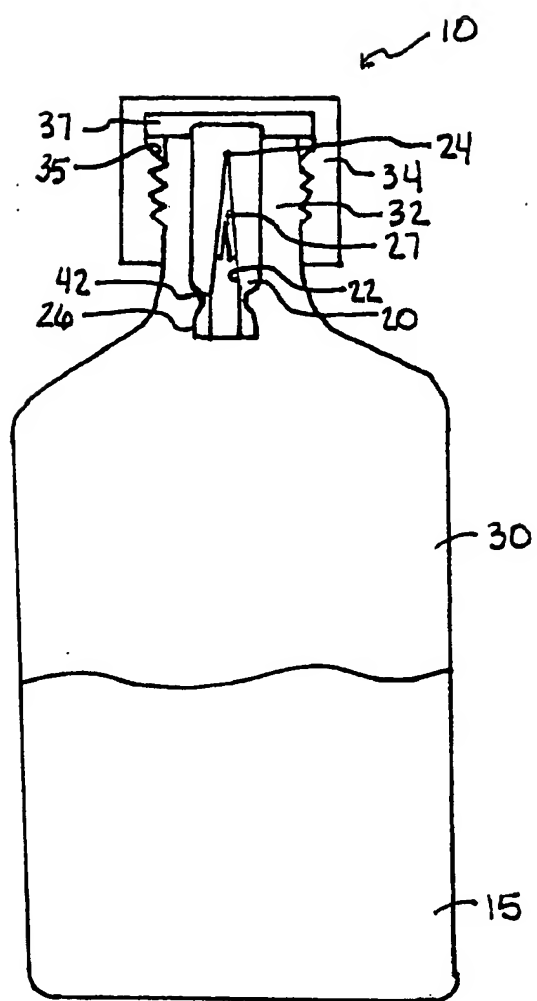


FIG. 1

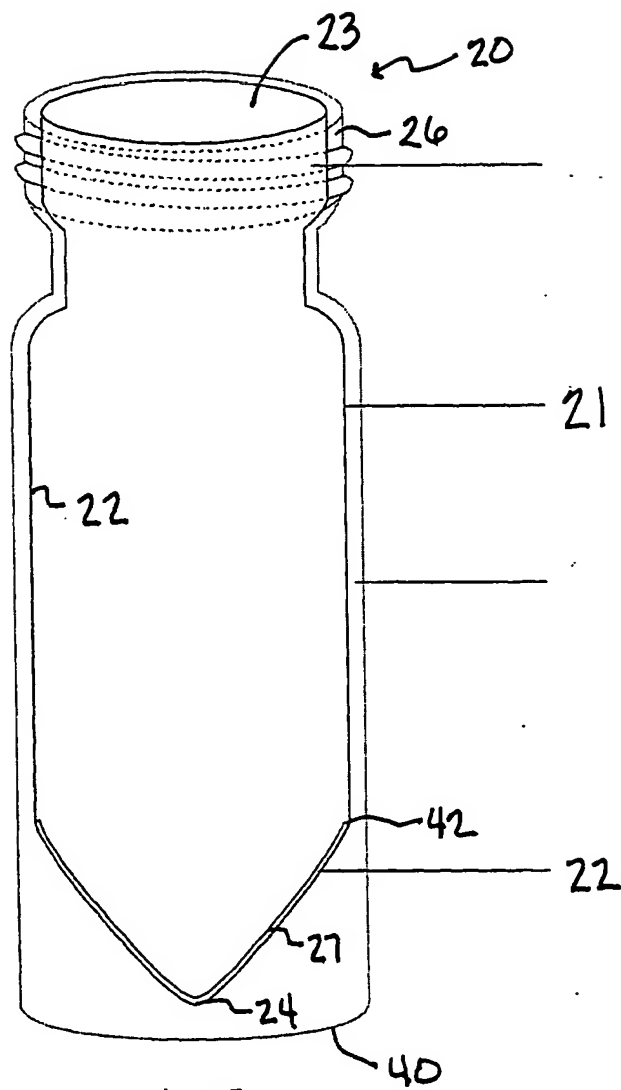


FIG. 2

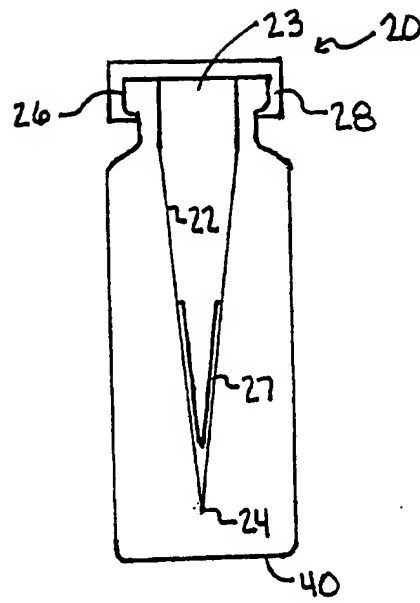


FIG. 3

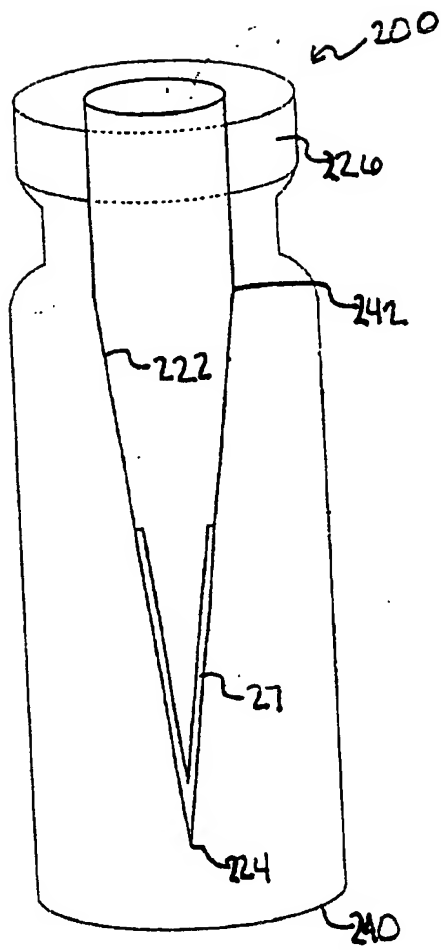


FIG. 4

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